(-)-ARMEPAVINE: GRADIENT-ENHANCED 2D NMR STUDY AND X-RAY CRYSTAL STRUCTURE DETERMINATION*

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(-)-Armepavine, a benzylisoquinoline alkaloid isolated from *Papaver caucasicum*, was examined by gradient-enhanced 2D NMR spectroscopy and X-ray diffraction analysis. Unambiguous ¹H, ¹³C, and ¹⁵N NMR assignment of the title alkaloid is reported.

Key words: (-)-Armepavine; Isoquinoline alkaloids; *Papaver caucasicum*; Gradient-enhanced 2D NMR spectroscopy; X-Ray crystallography.

(-)-Armepavine, (1*R*)-1-(4'-hydroxybenzyl)-6,7-dimethoxy-2-methyl-1,2,3,4-tetrahydroisoquinoline (1), was isolated for the first time from *Papaver armeniacum* (L.) DC. (*Papaveraceae*) by Konovalova *et al.*² in 1935 and its structure was solved by the same authors³.

(-)-Armepavine was later reported in *P. caucasicum* MARSCH.-BIEB.^{4,5} and in several other related *Papaver* species (*cf.* ref.⁶). Enantiomeric (+)-(*S*)-armepavine occurs in *Argemone turnerae* A.M. POWELL (*Papaveraceae*)⁷. (-)-(*R*)-Armepavine was also sporadically found in some plant species of the *Celastraceae*, *Magnoliaceae*, *Nymphaeaceae* (in the racemic form) and *Rhamnaceae* families. A goniometric study of armepavine crystals has been conducted by Uzbek authors⁸ in 1962. However, no relevant geometric parameters were reported. Steric correlation of (+)-*O*-methylarmepavine and (+)-laudanidine showed that the stereogenic carbon atom in natural (-)-armepavine has the *R* configuration⁹. It is known for a long time that in the benzylisoquinoline series, the levorotatory and dextrorotatory alkaloids have the *R* and *S* configurations, respectively^{10,11}.

^{*} Part XCVII in the series Alkaloids of the *Papaveraceae*; Part XCVI: see ref.¹.

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Recently, armepavine oxalate and methiodide have been tested on antiplatelet activity and stated that they exhibited weak but significant inhibition of platelet aggregation¹². In this paper, we report a detailed account of 2D NMR and crystal structure determination of the title alkaloid.



EXPERIMENTAL

(-)-Armepavine, m.p. 147–148 °C, $[\alpha]_{D}^{22}$ –119 ± 2° (*c* 0.5 M, CHCl₃) was isolated from *Papaver caucasicum* MARSCH.-BIEB. as described in ref.⁵. Colourless crystals were obtained from diethyl ether. ¹H, ¹³C, and 2D NMR spectra were recorded on a Bruker Avance DRX 500 (500/125 MHz) spectrometer in CDCl₃ at 303 K, δ values are in ppm. Signals were referenced to TMS as an internal standard. DQF-COSY (ref.¹³), NOESY (ref.¹⁴), HSQC (ref.¹⁵) and HMBC (refs^{16–18}) spectra were recorded using a 5 mm triple-resonance probe head equipped with a gradient coil with following parameters: DQF-COSY spectra: relaxation 2.2 s, gradient ratio 15 : 30 G/cm, MC2 States-TPPI; NOESY spectra: relaxation delay 2.3 s, mixing time 600–800 ms, MC2 States-TPPI; HSQC spectra: relaxation delay 2.3 s, delay for evolution of ¹H-¹³C coupling constants 3.33 ms (optimized for ¹J(H,C) coupling of 150 Hz), gradient ratio 12 : 36 : ±6 G/cm, GARP decoupling during acquisition, MC2 echo-antiecho; HMBC spectra: sequence¹⁸ D1-90°(¹H)-D6-90°(¹³C)-t₁/2-G1-D16-180°(¹H)-G2-D16-t₁/2-90° (¹³C)-G3-D16-ACQ(t₂), D16 = 100 µs, G = 1 ms, MC2 QF; ¹H-¹³C correlation: D1 = 2.7 s, D6 = 66.6 ms, G1 : G2 : G3 = 30 : 18 : 24 G/cm; ¹H-¹⁵N correlation: D1 = 3.5 s, D6 = 100 ms, G1 : G2 : G3 = 42 : 18 : 30 G/cm.

The diffraction data were collected on a KUMA KM-4 four-circle single crystal diffractometer using the ω -2 θ scan mode. The structure was solved by the direct method using SHELXS86 program¹⁹. All 24 non-hydrogen atoms were refined anisotropically by the weighted full-matrix least-squares procedure on F^2 with weight $w = 1/[\sigma^2(F_o^2) + (0.045P)^2 + 0.25P]$, where $P = (F_o^2 + 2F_c^2)/3$, using the SHELXL93 program package²⁰. All hydrogen atoms were localized from difference Fourier map and refined isotropically. Our choice of the *R* enantiomer was induced by the fact that we had the authentic (-)-*R*-armepavine isolated from plant material⁹⁻¹¹. The molecule was drawn by ORTEP (ref.²¹). Details about measurements and refinement are summarized in Table III.

RESULTS AND DISCUSSION

The ¹H and ¹³C chemical shifts are summarized in Table I. The standard ¹H NMR spectrum of (–)-armepavine^{22,23} in CDCl₃ displayed distinct singlets of the NMe, 6-OMe, and 7-OMe groups. The assignments of ¹H multiplets between 2.6 and 3.8 ppm was based on the gradient-enhanced phase-sensitive DQF-COSY experiment¹³ where clear interactions of H-1 with H-11x and H-11y were found. DQF-COSY also indicated

interactions between the H-2' (H-6') and H-3' (H-5') hydrogen atoms. There was also observed a correlation between H-3x and H-3y, H-4x and H-4y and the interaction of the H-3 signal with H-4. The NOESY experiment¹⁴ enabled us to distinguish between H-11x and H-11y, H-3x and H-3y, and to assign the signals of H-5, H-8, H-9, and H-10. The NOESY interactions of H-10 vs H-8 (C-10...H-8 2.50 Å) and H-9 vs H-5 (C-9...H-5 2.50 Å) were found. Considering the conformation in the solution and in the crystal (Fig. 1) the H-11x atom is close to H-8 (H-11x...H-8 3.54 Å vs H-11y...H-8 3.94 Å), H-11y reaches the proximity of NMe (H-11y...C-12 2.50 Å). A weak NOESY interaction of the H-3y signal with the H-11 atoms signal was also observed. The aromatic H-atoms resonated within the range of 5.9-6.9 ppm. The most up-field shifted is the H-8 atom (δ 5.98) which is near H-11x and probably under influence of the benzene ring anisotropy. Similarly, the 7-OMe group is slightly shifted (3.53 ppm) compared to 6-OMe (3.82 ppm). For an assignment of carbon atoms bound to the hydrogen atoms the gradient-enhanced HSQC pulse sequence¹⁵ was applied. The pulsed-field-gradient¹⁶⁻¹⁸ HMBC record led to the unambiguous assignment of quaternary carbon atoms. The signal of the H-9 hydrogen atoms showed a three-bond scalar interaction with C-6. On the other hand the H-atoms at C-10 correlate with the signal of C-7. Heteronuclear multiple bond interactions were also detected by gradient-enhanced phase-sensitive HSQC (GSQMBC, ref.²⁴). All observed scalar as well as dipolar interactions are displayed in Table II. The gradient-enhanced ¹H-¹⁵N HMBC (refs^{25,26}) spectrum indicated the interactions of H-12, H-11, H-3, and H-4 with the nitrogen atom N-2 at 32.70 ppm.

The X-ray analysis of (–)-armepavine (Tables III and IV, Fig. 1) showed that both the aromatic rings do not deviate significantly from planarity. The average deviation of the atoms C1'-C2'-C3'-C4'-C5'-C6' from the least square plane through the atoms²⁷ is 0.0047



FIG. 1 A perspective view of (-)-(R)-armepavine with atom numbering

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Å or 0.5 σ/δ , respectively, the average deviation of the atoms C4a–C5–C6–C7–C8–C8a from the plane is 0.0027 Å or 0.2 σ/δ , respectively. The angle between both the least square planes is 59.3(4)°. The adjacent oxygen atoms in the 6-OMe, 7-OMe, and 4'-OH groups are in the aromatic ring planes as indicated by corresponding torsion angles (Table IV). The hydrogenated part of isoquinoline adopts a distorted half-chair confor-

Atom	${}^{1}\mathrm{H}^{b}$	¹³ C	¹ <i>J</i> (H,C), H
1	3.72 (5.1,8.3)	64.81	137
3	-	45.93	136
3x	2.83 ^c	-	
3у	3.26 ^c	-	
4	_	24.51	127
4x	2.88^{c}	-	
4y	2.63 ^{<i>c</i>}	-	
4a	-	125.14	
5	6.56 s	111.23	156
6	-	147.33	
7	_	146.21	
8	5.98 s	111.27	156
8a	-	128.62	
9 (6-OMe)	3.82 s	55.70	144
10 (7-OMe)	3.53 s	55.43	144
11	-	40.26	127
11x	2.73 (8.3,13.6)	_	
11y	3.14 (5.1,13.6)	_	
12 (NMe)	2.51 s	41.95	134
1'	_	130.59	
2',6'	$6.89 d (8.5)^d$	130.69	156
3',5'	$6.64 d (8.5)^d$	115.38	157
4'	-	154.95	
4'-OH	5.40 br s		

TABLE I			
¹ H and ¹³ C NMR	data (δ,	ppm) of	(-)-armepavine ^a

^{*a*} For numbering the atoms, see Fig. 1. ^{*b*} Coupling constants (in Hz) are given in parentheses. ^{*c*} Coupling constants not determined due to complexity of signals. ^{*d*} A,A',X,X' spin system.

	1	7	3x	3y	4 x	4y	4a	S	9	٢	×	8a	6	10	11x	11y	12	1,	2′,6′	3',5'	, 4
-	c		p	р			р	р			b,d	р			a,b,d	a,b,d	b,d	р	b,d		
7		I	e	e	e										e	e	e				
3x	р	e	ပ	a,b,c	a,b,d	a,b,d	р										þ,d				
3y	р	e	a,b,c	ပ	a,b,d	a,b,d	р										b,d				
4 x		e	a,b,d	a,b,d	с	a,b,c	p	b,d			p	р									
4y			a,b,d	a,b,d	a,b,c	c	р	b,d			р	р									
4a	р		q	p	q	p	I	p			p										
5	q				b,d	b,d	q	c	q	p	q	q	q								
9								p	I	Ι	p		р								
٢								р	I	Ι	р			р							
8	b,d				р	р	р	р	р	р	ပ	р		q	q				q		
8a	р				р	р		р			р	I			р	p					
6								q	р				ပ								
10										р	q			ပ					q		
11x	a,b,d	e									q	p			c	с		р	p		
11y	a,b,d	e										p			c	c		р	р		
12	b,d	e	b,d	b,d													c				
1,	p														p	p		Ι	р	p	
2′,6′	b,d										q			q	р	р		р	c	a,d	р
3',5'																		р	a,d	ပ	р
, 4																			p	p	Ι

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mation with the C-3, N-2, and C-4 atoms significantly deviated from the best mean plane of isoquinoline by 0.309(3) Å, 0.448(2) Å, and 0.44(3) Å, respectively, as in other 1,2,3,4-tetrahydroisoquinoline systems^{28,29}. Puckering parameters²⁷ of the C1–N2–C3–C4–C4a–C8a heterocyclic ring are Q = 0.511(14) Å, $\phi = 88(2)^{\circ}$ and $\theta =$ 48.5(12)°. Armepavine crystallized with one molecule of water which is bound by a hydrogen bond to the phenolic hydroxy group. The length of the hydrogen bond H-15...O-16 is 1.75(4) Å. The O-15...O-16 distance is 2.65(3) Å. The mean value of the three C–N–C bond angles is 112.4(2)° indicating an sp³ hybridization of the N atom.

TABLE III

Diffractometer	KUMA KM-4
Formula, m.w.	C ₁₉ H ₂₃ NO ₃ . H ₂ O, 331.4
Temperature	298(2) K
Wavelength	0.71073 Å
Crystal system	orthorhombic
Space group	<i>P</i> 2 ₁ 2 ₁ 2 ₁
Unit cell dimensions	a = 6.2909(4) Å, $b = 10.822(2)$ Å, $c = 25.823(6)$ Å
Volume	1 758.0(5) Å ³
Z	4
Density (calculated)	1.252 Mg/m ³
Absorption coefficient	0.087 mm^{-1}
<i>F</i> (000)	712
Crystal size	$0.70\times0.60\times0.50~\mathrm{mm}$
θ range for data collection	2.04–26.57°
Index ranges	$-7 \le h \le 1, \ 0 \le k \ \le 13, \ 0 \le l \le 32$
Reflections collected	2 189
Independent reflections	2 157 $[R(int) = 0.0090]$
Refinement method	full matrix least-squares on F^2
Data; restraints; parameters	2 157; 0; 317
Goodness-off-fit on F^2	0.975
Final <i>R</i> indices $[I > 2\sigma(I)]$	R1 = 0.0324, wR2 = 0.0790
<i>R</i> indices (all data)	R1 = 0.0441, wR2 = 0.0849
Largest different peak and hole	0.150 and –0.127 e ${\rm \AA}^{-3}$

Crystal data and structure refinement for (-)-armepavine

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1 1		•
(-)	- Arm	enavine
v	/ 1 1111	cpavine

TABLE IV

Bond lengths (Å), bond angles and selected dihedral angles (°) in (–)-armepavine

Atoms	Distances	Atoms	Distances
C1–N2	1.478(3)	C7–O14	1.376(2)
C1–C8a	1.514(3)	C8–C8a	1.405(3)
C1-C11	1.546(3)	C9-013	1.430(3)
N2-C12	1.464(3)	C10-O14	1.433(3)
N2-C3	1.464(4)	C11-C1'	1.499(4)
C3–C4	1.507(4)	O15-C4′	1.360(3)
C4–C4a	1.505(3)	C1′–C6′	1.377(4)
C4a–C8a	1.384(3)	C1'-C2'	1.402(4)
C4a–C5	1.393(3)	C2'–C3'	1.371(4)
C5–C6	1.374(3)	C3'-C4'	1.384(4)
C6013	1.364(2)	C4′–C5′	1.381(4)
C6–C7	1.403(3)	C5′–C6′	1.377(4)
C7–C8	1.371(3)		
Atoms	Angles	Atoms	Angles
N2-C1-C8a	109.5(2)	C7–C8–C8a	121.3(2)
N2-C1-C11	114.5(2)	C4a–C8a–C8	118.7(2)
C8a-C1-C11	112.4(2)	C4a-C8a-C1	122.1(2)
C12-N2-C3	111.5(2)	C8-C8a-C1	119.2(2)
C12-N2-C1	113.5(2)	C1′-C11-C1	113.6(2)
C3-N2-C1	112.1(2)	C6013C9	116.1(2)
N2-C3-C4	109.3(2)	C7-O14-C10	117.2(2)
C4a-C4-C3	111.5(2)	C6'-C1'-C2'	116.7(2)
C8a–C4a–C5	119.7(2)	C6'-C1'-C11	121.7(2)
C8a–C4a–C4	121.3(2)	C2'-C1'-C11	121.6(2)
C5-C4a-C4	119.0(2)	C3'-C2'-C1'	121.3(3)
C6–C5–C4a	121.6(2)	C2'-C3'-C4'	120.9(3)
O13-C6-C5	125.3(2)	O15-C4'-C5'	123.2(3)
O13-C6-C7	115.9(2)	O15-C4'-C3'	118.3(2)
C5-C6-C7	118.8(2)	C5'-C4'-C3'	118.5(3)
C8–C7–O14	124.7(2)	C6'-C5'-C4'	120.2(3)
C8–C7–C6	119.9(2)	C5'-C6'-C1'	122.5(2)
O14–C7–C6	115.4(2)		
Atoms	Dihedral angles	Atoms	Dihedral angles
C4a-C5-C6-O13	178.0(14)	O14-C7-C8-C8a	-179.9(13)
C5-C6-C7-O14	179.9(13)	C2'-C3'-C4'-O15	178(2)
O13-C6-C7-O14	1(2)	O15-C4'-C5'-C6'	-179(2)
013-C6-C7-C8	-177.5(13)		

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REFERENCES

- 1. Slavik J., Slavikova L.: Collect. Czech. Chem. Commun. 61, 1815 (1996).
- 2. Konovalova R., Yunusov S., Orechov A.: Ber. Dtsch. Chem. Ges. 68, 2158 (1935).
- 3. Yunusov S., Konovalova R. A., Orechov A. I.: Zh. Obshch. Khim. 10, 641 (1940).
- 4. Kuhn L., Pfeifer S., Slavik J., Appelt J.: Naturwissenschaften 51, 556 (1964).
- 5. Preininger V., Appelt J., Slavikova L., Slavik J.: Collect. Czech. Chem. Commun. 32, 2682 (1967).
- 6. Preininger V. in: The Alkaloids (A. Brossi, Ed.), Vol. 29, p. 1. Academic Press, San Diego 1986.
- 7. Stermitz F. R., Kim D. K., Larson K. A.: Phytochemistry 12, 1355 (1973).
- 8. Parpiev N. A., Tashpulatov Yu.: Dokl. Akad. Nauk Uzb. SSR 19, 17 (1962).
- 9. Ferrari C., Deulofeu V.: Tetrahedron 18, 419 (1962).
- 10. Santavy F. in: *The Alkaloids* (R. H. F. Manske, Ed.), Vol. 12, p. 333. Academic Press, New York 1970.
- 11. Santavy F. in: *The Alkaloids* (R. H. F. Manske and R. Rodrigo, Eds), Vol. 17, p. 385. Academic Press, New York 1979.
- 12. Chen K. S., Ko F. N., Teng Ch. M., Wu Y. Ch.: J. Nat. Prod. 59, 531 (1996).
- 13. Bodenhausen G., Freeman R., Turner D. L.: J. Magn. Reson. 27, 511 (1977).
- 14. Jeener J., Meier B. H., Bachmann P., Ernst R. R.: J. Chem. Phys. 71, 4546 (1979).
- 15. Bodenhausen G., Ruben D. J.: Chem. Phys. Lett. 69, 185 (1980).
- 16. Bax A., Summers M. F.: J. Am. Chem. Soc. 108, 2093 (1986).
- 17. Hurd R. E.: J. Magn. Reson. 87, 422 (1990).
- 18. Marek R., Sklenar V., Dostal J., Slavik J.: Tetrahedron Lett. 37, 1655 (1996).
- Sheldrick G. M.: SHELXS86. Program for Crystal Structure Solution. University of Gottingen, Gottingen 1986.
- Sheldrick G. M.: SHELXL93. Program for Crystal Structure Refinement from Diffraction Data. University of Gottingen, Gottingen 1993.
- Johnson C. K.: ORTEP. Report ORNL-3794. Oak Ridge National Laboratory, Oak Ridge, TN 1965.
- 22. Torres R., Monache F. D., Bettolo G. B. M.: J. Nat. Prod. 42, 430 (1979).
- 23. Bishay D. W., Kowalevski Z., Phillipson J. D.: Phytochemistry 12, 693 (1973).
- 24. Marek R., Kralik L., Sklenar V.: Tetrahedron Lett. 38, 665 (1997).
- 25. Martin G. E., Crouch R. C., Sharaf M. H. M., Schiff P. L., Jr.: J. Nat. Prod. 59, 2 (1996).
- 26. Marek R., Dostal J., Slavik J., Sklenar V.: Molecules 1, 166 (1996).
- Nardelli N.: PARST95. System of Computer Routines for Calculating Molecular Parameters from the Results of Crystal Structure Analyses. University of Parma, Parma 1995.
- 28. Fridrichsons J., Mathieson A. M.: Tetrahedron 24, 5785 (1968).
- Dokurno P., Gdaniec M., Kosturkiewicz Z., Matecka D., Rozwadowska M. D.: Acta Crystallogr., C 49, 517 (1993).